

# POLLEN GRAINS TAXONOMY MATCHES DNA-BARCODING TAXONOMY IN DISCRIMINATION BETWEEN SOME CONVOLVULACEAE TAXA

Hany S. Abd El-Raouf<sup>1,2</sup>, Mohamed Helmy<sup>3\*</sup>, Mohamed Awad<sup>4</sup> and Ahmed M. El-Taher<sup>1</sup>

<sup>1</sup> Agricultural Botany Department, Faculty of Agriculture, Al-Azhar University, Cairo, Egypt
 <sup>2</sup> Biology Department, University College, Taif University, Turaba, Kingdom of Saudi Arabia
 <sup>3</sup> Singapore Institute of Food and Biotechnology Innovation (SIFBI), Agency for Science,

Technology and Research (A\*STAR), Singapore.

<sup>4</sup> Department of Biotechnology, Faculty of Agriculture, Al-Azhar University, Cairo, Egypt \* Corresponding author: mohamed\_helmy@sifbi.a-star.edu.sg

## Abstract

Palynological investigation relays on morphology and seed coat characters. It's a well-known approach to be used in discrimination between related plants. While the DNS barcoding used specific regions in the plant genome to identify the plant species/sub-species. Here, we compared pollen grain taxonomy and DNA barcoding taxonomy of 12 taxa belonging to the *Convolvulaceae* family growing in Egypt. The pollen grains were examined by the scanning electron microscope (SEM) then their features were recorded in a data matrix to compare their characters and create a phylogenetic tree. The gene sequences of the MatK gene, that is wildly used in the DNA barcoding, of the same 12 plants were compared to build another phylogenetic tree based on the sequence similarity between the gene sequences. Our results show high similarity between the two phylogenetic trees indicating the high efficiency of the pollen grain taxonomy and a promising integration of micro-morphological approaches and molecular approaches in discrimination between plants.

Keywords: Pollen grains taxonomy, DNA barcoding, Argyria, Convolvulus, Ipomoea, Merremia.

## Introduction

A palynological investigation is a popular tool in plant taxonomy, especially with the invention of the highresolution power microscopes as the scanning electron microscopes (SEM). The basis of this branch is Erdmann's 1952 handbook of palynology and its second edition by Nilson and Praglowski (Erdtman, 1952). On the other hand, the modern systematics communities depend on genetic bases (Wen et al., 2017). DNA barcode is the most popular DNA-based method for species identification and taxonomy clarification (Hollingsworth et al., 2011). Several chloroplast genes, intragenic and intergenic regions are used as a plant DNA barcodes such as trnfM-trnT, trnH-psbA, trnL(p6) and ycf1 (Awad et al., 2017; Choi et al., 2015; Kress et al., 2005) The consortium for the barcode of life adopted MatK and rbcL chloroplast genes as core genes for plant taxonomy, identification and systematics (Hollingsworth et al., 2009).

The taxonomic significance of pollen morphology in Convolvulaceae has long been recognized. Hallier (1893) was the first to divide this family based on pollen features into two groups, Echinoceniae and Psiloconiae based on their echinate or psilate exine, respectively (Hallier, 1893). The genus Convolvulus was put in Psiloconiae in which the pollen has either a psilate or a granulated surface. In the division of Gamble 1923, the family was divided into two groups based on echinate and non-echinate pollen grains with the genus Convolvulus included in the latter one (Gamble, 1923). Erdtman 1952 separated the Convolvulacious pollen grains into two groups. Namely Ipomoea type and other types, in which he includes genus Convolvulus (Erdtman, 1952). The Ipomoea type possesses pollen grains which are polyporate with a thick exine and are echinate, whereas the Convolvulus type is distinctly perforate. O. Donell 1955 separated genus Convolvulus and Calystegiaon the basis of Halliers aperture descriptions, but he reported that in *Convolvulus* the pollen is 3-colporate (Õ Donell, 1955).

Sengupta 1972 carried out a comprehensive study in the family and divided it into four main pollen types based on the number and distribution of apertures (Sengupta, 1972). Pollen characters provide taxonomically valuable information that has long been used in the classification of the Convolvulaceae. Indeed, many keys require knowledge of the pollen surface features and apertures to arrive at a genus name (Austin et al., 1998). According to Boulos 2000, 12 Convolvulaceae species are growing in Egypt (Boulos, 1999). In this article are using pollen characters and the sequence of MatK gene for the 12 species which grow in Egypt to discrimination these species and compare the pollen characters and the DNA barcoding results.

## **Materials and Methods**

#### **Taxa acquisition**

Twelve taxa of family *Convolvulaceae*, some species of genus *Convolvulus* collected from Marsa Matruh, a resort town on Egypt's Mediterranean coast, and the other taxa were collected from the Herbarium of Orman garden (Giza governorate, Egypt) through 2014 and 2016 (Table.1). The identification of the collected specimens was achieved by comparing their morphological characters with the characters of the previously identified plants as published by (Boulos, 1999).

#### **Pollen grains imaging**

Pollen grains were separated from anthers, for each taxon three specimens were used, and from each specimen at least five anthers were examined. For Scanning Electron Microscope (SEM) examination, pollen grains were coated by a gold sputter coater (Sp1-Module), then examined by SEM (JEOL JSM-5500LV) JEOL Ltd., Japan, by using low vacuum mode at a magnification of 1200-1600x at the Regional Center of Mycology and Biotechnology, Cairo, Egypt. We used the terminology of Punt *et al.* 2007 and

Sugandha to describe the morphological characters (Punt *et al.*, 2007).

## **Bioinformatics and data analysis**

Multi-Variate Statistical Package Programme (MVSP) was used to analyze the data of pollen grains characters (Sneath, 1973). Twelve MatK sequences were retrieved from **NCBI** for the species (Table 12 1). TTCATTACTCKAAAGAGGTC and AADATTTCTGYA TATACG manually designed and used as a forward and reverse primers respectively, for In-silico PCR. A sequence length of 675 bp from the MatK gene was used for multiple sequence alignment and maximum likelihood (ML) phylogenetic tree construction using CLC v.7 (CLC bio, Aarhus, Denmark) with 100 bootstrap replicates. We used iTol v.5 for tree visualization (Letunic and Bork, 2019). The trees were redrawn for clarity and the original trees are included in the supplementary files for references (Figure S1).

## **Results and Discussion**

## Palynological investigation

The pollen taxa exhibited a wide range of size variation. All investigated taxa of three genera *Argyria*, *Ipomoea* and *Merremia* are large (50-100um) while genus *Convolvulus* is medium (25-50um). All taxa are isopolar except *C. arvensis* (Figure 1-c) and *C. oleifolius* (Figure 1-f) are apolar. All taxa show radically symmetrical as in *Argyria*, *Ipomoea* and two species belonging to *Convolvulus* genus (*C. dorycnium* and *C. pilosellifolius*) (Figure 1-d and g) while the other species are bilateral such as (*C. althaeoides*, *C. arvensis*, *C. lineatus* and *C. oleifolius* (Figure 1-b, c and f). The examined taxa were separated into two groups, a group with colpate aperture (tricolpate) included genus *Convolvulus* and *Merremia*, (Figure 1-b, c, d, e, f, g and i) and the other group with porate (polyporate) included *Argyria* and *Ipomoea* (Figure 1-a, h, i, j and k).

According to the ratio between polar axis and equatorial diameter in the equatorial view determined by the shape of pollen grains, most of the examined taxa are spheroidal shapes such as Argyria nervosa, Ipomoea stolonfera and Merremia dissecta (Figure 1-a, k, and l), two taxa only have prolate shape, Convolvulus arvensis and C. oleifolius (Figure 1-c and f) while C. althaeoides only has prolate-spheroidal (Figure 1-b) and Ipomoea cairica has oblate shape (Figure 1h). Sculpturing is granulate in all species of Convolvulus and Merremia dissecta, (Figure. 1-b, c, d, e, f, g and i) while it is echinate in Argyria nervosa and species of Ipomoea (Figure. 1-a, h, i, j and k). Polar axis length varies from 35µm. to 91µm. Equatorial diameter varies from 20µm. to 91µm. while aperture length varies from 5µm. to 35µm. All observed palynological characters (both quantitative and qualitative) of 12 investigated taxa belonging to family Convolvulaceae are listed in Table 1 and the SEM images are shown in Figure 1.



**Fig. 1 :** SEM images of pollen grains of the 12 taxa of the family *Convolvulaceae*. A) *Argyria nervosa, B) Convolvulus* althaeoides, C) C. arvensis, D) C. dorycnium, E) C. lineatus, F) C. oleifolius, G) C. pilosellifolius H) Ipomoea cairica, I) I. eriocarpa, J) I. hederacea K) I. stolonfera and L) Merremia dissecta







Fig. 2-B





**Fig. S1 :** Original phylogenetic trees. A) Dendrogram is constructed on the basis of 12 pollen grains features. B) The phylogenetic tree resulted from the MatK gene sequence analysis.

These results are in agreement with the known features of pollen grains of Convolvulus arvensis (medium size, prolate shape, and tricolpate) (Ashfaq et al., 2018). Also, it is in agreement with what was reported about Ipomoea cairica in size and sculpturing but contrary to wat reported about its shape, which was reported as subspheroidal (V et al., 2011). The four species belonging to genus Ipomoea are in agreement with the present work in the type of sculpturing and the present work in features of Argyreia nervosa in symmetry, polarity, aperture type, shape and sculpturing (Saensouk, S. and Saensouk, 2018).

#### **Cluster analysis**

Cluster analysis produces a phylogenetic tree of 12 taxa, based on the data matrix (Table 1). The resultant phylogenetic tree is shown in Figure 2A which shows that the studied taxa are divided into two groups. The first group includes all species belonging to genus Convolvulus plus Merremia dissecta which showed clear similarity with Convolvulus except in the pollen grain size. Species of Convolvulus differed in polarity, similarity, the shape of pollen grains in addition to polar axis and equatorial axis as shown in Table 1. The second group includes both Argyria nervosa and Ipomoea species and we depend on statics characters to discriminate between Ipomoea species and Argyria. Both were similar in polarity, symmetry, apertures type, number of aperture and exine sculpturing. The present numerical analysis results were in harmony with those obtained before on some species of fabaceous plants (Khattab and El-Kobisy, 2007).

#### **DNA-barcoding investigation**

To investigate the agreement between palynological taxonomy and genomic taxonomy in Convolvulaceae members discrimination we compared the MatK gene sequence in the 12 taxa. MatK is a chloroplast gene that is widely used for taxonomical and species identification purposes. A representative MatK sequence was retrieved from NCBI for each species. Then, we designed a forward and a reverse degenerative primers (see methods) to unify the sequence borders to avoid the sequence bias coming from unaligned nucleotides in flanks. In total, 675 pb used for ML gene tree construction. MatK phylogenetic tree showed a harmonious taxonomical topology with those palynological taxonomy approaches (Figure 2B).

Convolvulus species distinguished in a separate clade with a sister lineage *Merremia dissecta*, both shared the same ancestral node. This clade agreed with colpate operture clade in palynological dendrogram (Figure 2B). On the other hand, *Ipomoea* species showed a gradual lineage development from I. imperati (stolonfera), I. cairica and I. hederacea, respectively, then I. eriocarpa and Argyria nervosa as a sister lineages from the same internal node (Figure 2B). The overall figure agreed with porate clade in palynological dendrogram by grouping Argyria nervosa with Ipomoea species. Moreover, this agreed with the modern taxonomy (NCBI:txid139741). The only difference is that the Palynological dendogram shows that Argyria nervosa has an internal node with I. hederacea, while the MatK phylogenetic tree and Pollen grain identification key showed I. eriocarpa is closer to Argyria nervosa than I. hederacea. This also agreed with the whole chloroplast phylogenetic tree (Eserman et al., 2014).

## Identification key of the 12 Convolvulaceae family taxa based on the observed pollen grains features

l -	Pollen grains: tricolpate, granulate a-pollen grains, large size (50-100) µm, spheroidal Merremia dissecta
	b-pollen grains, medium size (25- 50)µm
	bb- pollen grains, bilateral
	bb1- pollen grains, prolate-spheroidal C. althaeoides
	bb2- pollen grains, spheroidal C. lineatus
	<ul> <li>c- pollen grains, radially symmetrical, spheroidal shape</li> <li>c1- polar axis 40um</li></ul>
	d- pollen grains, isopolar, prolate shape
	d1- polar axis 45um
	d2- polar axis 30um
)_	Pollen grains polyporate, echinate
-	A pollen grains, spheroidal shape
	THE DOTTOM MEMORY OPPING OF THE DESCRIPTION OF THE

A1- polar axis 90um., aperture length 7.5um. ... Argyria nervosa A2- polar axis 84um., aperture length 6um. ..... I. eriocarpa A3- polar axis 89um., aperture length ..... I. hederacea A4- polar axis 91um., aperture length 7.7um. .. I. stolonfera

#### Conclusion

We investigated the taxonomy of all the members of the Convolvulaceae family that grow in Egypt, 12 taxa in total. We used palynological taxonomy based on pollen grain characters observed through SEM imaging and genomic taxonomy through the analysis of the MatK chloroplast gene sequence. We built two phylogenetic trees based on the two analyses and compare them to investigated the agreement between the two approaches and we created an identification key for the discrimination between the 12 taxa based on the palynological analysis. Our results show that the 12 taxa are divided into two groups, a group includes all species belonging to genus Convolvulus plus Merremia dissecta and another group includes both Argyria nervosa and Ipomoea species. Furthermore, our results show a harmony between the palynological and genomic approaches as both of them gave similar results.

B. pollen grain, oblate shape, polar axis 40um. aperture length 5um. .... I. cairica

**Table 1 :** Data matrix and accession numbers of the observed characters in the different taxa. The data matrix based on 11 characters of 12 taxa.

	Taxa	Accession Number	Polar axis µm	Equatorial axis µm	P/е µm	Shape of pollen grain*	Polarity Apolar+/isopolar-	Symmetry Radially symmetrical+/bilaterial-	Size Large+/medum-	Number of aperture Tri+/poly-	Apertural type Colpate+/porate-	aperture length µm	Exine sculpturing Granulate+/echinate-
1	Argyria nervosa	KR024898	90	90	90/90	S	+	+	+	-	-	7.5	-
2	Convolvulus althaeoides	KC529015	45	42	45/42	P-S	+	-	I	+	+	18	+
3	C. arvensis	KC529020	45	28	45/28	Р	-	-	-	+	+	35	+
4	C. dorycnium	KC529040	40	40	40/40	S	+	+	-	+	+	20	+
5	C. lineatus	KC529081	40	37	40/37	S	+	-	-	+	+	12	+
6	C. oleifolius	KC529089	30	20	30/20	Р	-	-	-	+	+	12	+
7	C. pilosellifolius	KC529016	35	35	35/35	S	+	+	-	+	+	12	+
8	Ipomoea cairica	FJ795793	40	60	40/60	0	+	+	+	-	-	5	-
9	I. eriocarpa	KF242483	84	84	84/84	S	+	+	+	-	-	6	-
10	I. hederacea	MG973747	89	89	89/89	S	+	+	+	-	-	7	-
11	I. stolonfera	MF064347	91	91	91/91	S	+	+	+	-	-	7.7	-
12	Merremia dissecta	KR024956	68	68	68/68	S	+	+	+	+	+	5.5	+

\* S: spheroidal, P-S: prolate-spheroidal, P: prolate, O: oblate)

#### References

- Ashfaq, S.; Zafar, M.; Ahmad, M.; Sultana, S.; Bahadur, S.; Khan, A. and Shah, A. (2018). Microscopic investigations of palynological features of convolvulaceous species from arid zone of Pakistan. Microsc. Res. Tech. 81: 228–239.
- Austin, D.; Mathew, P. and Sivadasan, M.T. (1998). Parallel and convergent evolution in the Convolvulaceae, 201-234.
- Awad, M.; Fahmy, R.M.; Mosa, K.A.; Helmy, M. and El-Feky, F.A. (2017). Identification of effective DNA barcodes for Triticum plants through chloroplast genome-wide analysis. Comput. Biol. Chem. 71.
- Boulos, L. (1999). Flora of Egypt. Al Hadara Pub.
- Choi, D.H.; Kwon, Y.M.; Kwon, K.K. and Kim, S.-J. (2015). Complete genome sequence of Novosphingobium pentaromativorans US6-1(T). Stand. Genomic Sci., 10: 107.
- Erdtman, G. (1952). Pollen morphology and plant taxonomy. GFF. https://doi.org/10.1080/11035895209453507
- Eserman, L.A.; Tiley, G.P.; Jarret, R.L.; Leebens-Mack, J.H. and Miller, R.E. (2014). Phylogenetics and diversification of morning glories (tribe Ipomoeeae, Convolvulaceae) based on whole plastome sequences. Am. J. Bot. 101: 92–103.
- Gamble, J.S. (1923). Flora of the Presidency of Madras. Antiquariaat A. Kok & Zn. B.V., Amsterdam, Netherlands.
- Hallier, H. (1893). Biodiversity and taxonomy of morphologischer and anatomischer. Grundlage. Bot. Jahrb.
- Hollingsworth, P.M.; Forrest, L.L.; Spouge, J.L.; Hajibabaei, M.; Ratnasingham, S.; van der Bank, M.; Chase, M.W.; Cowan, R.S.; Erickson, D.L.; Fazekas, A.J.; Graham, S.W.; James, K.E.; Kim, K.J.; John Kress, W.; Schneider, H.; van AlphenStahl, J.; Barrett, S.C.H.; van

den Berg, C.; Bogarin, D.; Burgess, K.S.; Cameron, K.M.; Carine, M.; Chacón, J.; Clark, A.; Clarkson, J.J.; Conrad, F.; Devey, D.S.; Ford, C.S.; Hedderson, T.A.J.; Hollingsworth, M.L.; Husband, B.C.; Kelly, L.J.; Kesanakurti, P.R.; Kim, J.S.; Kim, Y.D.; Lahaye, R.; Lee, H.L.; Long, D.G.; Madriñán, S.; Maurin, O.; Meusnier, I.; Newmaster, S.G.; Park, C.W.; Percy, D.M.; Petersen, G.; Richardson, J.E.; Salazar, G.A.; Savolainen, V.; Seberg, O.; Wilkinson, M.J.; Yi, D.K. and Little, D.P. (2009). A DNA barcode for land plants. Proc. Natl. Acad. Sci., U.S.A. 106: 12794–12797.

- Hollingsworth, P.M.; Graham, S.W. and Little, D.P. (2011). Choosing and Using a Plant DNA Barcode. PLoS One 6: e19254.
- Kress, W.J.; Wurdack, K.J.; Zimmer, E.A.; Weigt, L.A. and Janzen, D.H. (2005). Use of DNA barcodes to identify flowering plants. Proc. Natl. Acad. Sci. U.S.A. 102: 8369–8374.
- Letunic, I. and Bork, P. (2019). Interactive Tree of Life (iTOL) v4: recent updates and new developments. Nucleic Acids Res. 47, W256–W259. https://doi.org/10.1093/nar/gkz239
- Õ Donell, C. (1955). Convolvulaceae Argentinas. Lilloa, palynology-1, Angiosperm. Almqvist and Wiksell, Stockholm.
- Punt, W.; Hoen, P.P.; Blackmore, S.; Nilsson, S. and Le Thomas, A. (2007). Glossary of pollen and spore terminology. Rev. Palaeobot. Palynol. 143: 1–81.
- Saensouk, S. and Saensouk, P. (2018). Palynology of family Convolvulaceae in Thailand. Res. Knowl. 4: 16–33.
- Sengupta, S. (1972). On the pollen morphology of convolvulaceae with special reference to taxonomy. Rev. Palaeobot. Palynol. 13: 157–212.
- Sneath, P.H. and S.R.R. (1973). PesticidNumerical Taxonomy: The Principles and Practice of Numerical Classificationes Exposure: The Case of Workers on Growing Grapes in San Francisco Valley,

Petrolina/Brazil, 1st ed, Agricultural Sciences. W. H. Freeman, , San Francisco.

- V, R.A.; A, T.J. and Patil, G.V. (2011). Studies on pollen morphology of Ipomoea species (Convolvulaceae). Res. Plant Biol., 1: 41–47.
- Wen, J.; Harris, A.; Ickert-Bond, S.M.; Dikow, R.; Wurdack, K. and Zimmer, E.A. (2017). Developing integrative

systematics in the informatics and genomic era, and calling for a global Biodiversity Cyberbank. J. Syst. Evol. 55: 308–321.

Khattab, O.S. and El–Kobisy, K.H.S.E. (2007). Botanical studies on some genera of Mimosaceae and Caesalpiniaceae. II- Seed features. J. Agric. Sci. Mansoura Univ. 32.